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Science and Technology

# DESIGNING BIOCATALYSTS BASED ON ENZYMES CO-IMMOBILIZATION WITH APPLICATION FOR CASCADE REACTIONS

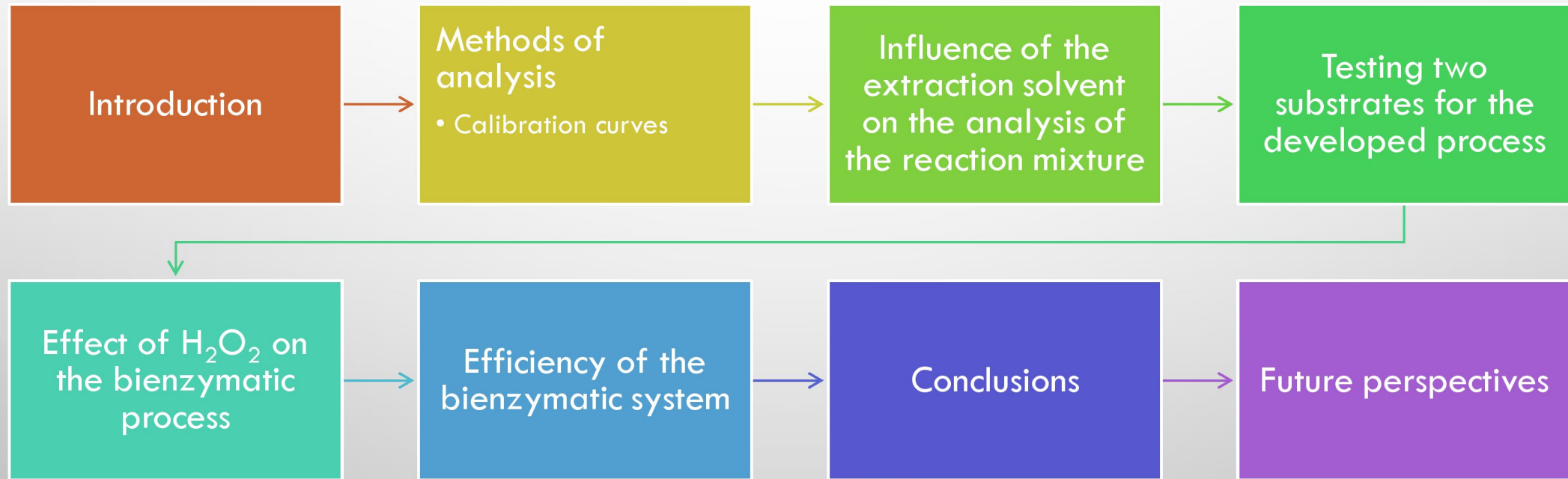
## Experimental report (I)

MASTER: CHEMISTRY OF ADVANCED MATERIALS

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COORDINATOR: ASSOC PROF. DR. MĂDĂLINA SĂNDULESCU

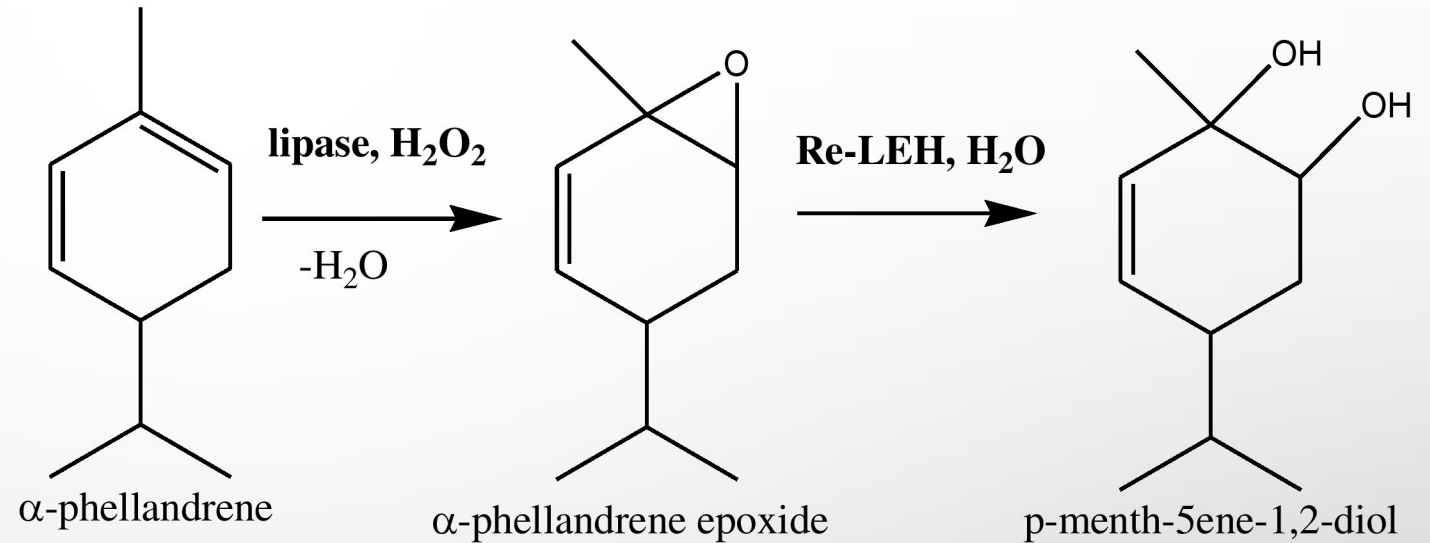
# OUTLINE



# Introduction

We propose a bienzymatic cascade system based on:

- Monoterpenoid epoxidation indirectly biocatalyzed by lipase from *Aspergillus niger*
- Epoxide hydrolysis directly biocatalyzed by hydrolase Re-LEH



In this experimental report, we present our results for:

▪ Analysis of reaction mixture

▪ Testing two substrates for the developed process

▪ Optimization of  $\text{H}_2\text{O}_2$  concentration

# Methods of analysis

## A sample contains:

- 1.6 M octanoic acid
- 1.6 M substrate
- 0.1 M PBS (pH 8)
- 4.39% v/v lipase from *Aspergillus niger*
- 2.81% v/v hydrolase Re-LEH
- > 0.44 mM H<sub>2</sub>O<sub>2</sub>

- contains: NaCl, KCl, Na<sub>2</sub>HPO<sub>3</sub>, K<sub>2</sub>HPO<sub>3</sub> in water - pH adjusted with NaOH  
 - is used to maintain a constant volume among all samples

## GC-FID (flame ionization):

- Requested the acetylation step for analysis
- Enantioseparation

Process enantioselectivity

## GC-MS (mass spectrometer):

- Does not need the acetylation step
- Reverse phase separation

## Sample analysis:

- Gas chromatography

24 h  
25°C  
1000 rpm  
(Thermoshaker)

## Extraction:

- 1:1 volume ratio in ethyl acetate or n-pentane

Agitation at room temperature (30 min)

24 h  
25°C  
1000 rpm  
(Thermoshaker)

## Acetylation:

- 100 µL sample
- 150 µL pyridine
- 66 µL acetic anhydride



## Masks unreacted substrate

Pre-treatment	Unreacted $\alpha$ -phellandrene (mols/L)
Extraction	$19.00 \times 10^{-2}$
Extraction + acetylation	$6.54 \times 10^{-2}$

Values obtained from interpolation on calibration curve

Substrate conversion

# Methods of analysis

## Calibration curves:

- For a proper determination of substrate and interested product in the reacted mixture

- 8 samples
- Starting with around 3.3 M the concentrations are halved till 0.05 M
- The final sample has 0 M
- Solvents : n-pentane and ethyl acetate

## Acetylation:

- 100  $\mu\text{L}$  sample
- 150  $\mu\text{L}$  pyridine
- 66  $\mu\text{L}$  acetic anhydride

24 h  
25°C  
1000 rpm  
(Thermoshaker)

## GC-MS:

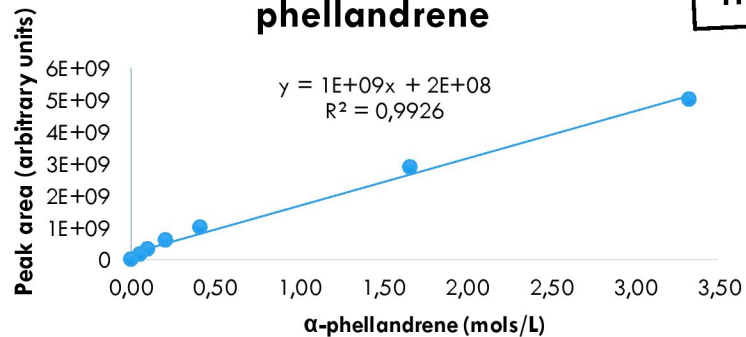
- More sensible in changes in substrate concentration

## GC-FID:

- Can distinguish between R and S isomers of resulting diols

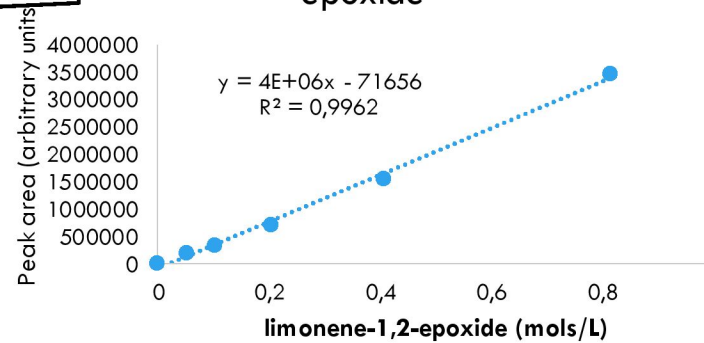
ethyl acetate

Calibration curve for  $\alpha$ -phellandrene

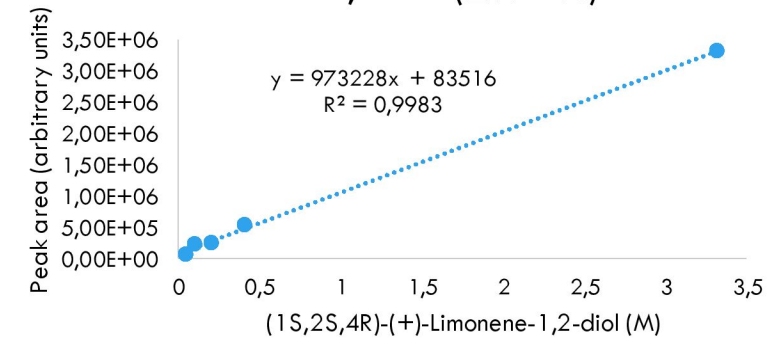


n-pentane

Calibration curve for limonene-1,2-epoxide



Calibration curve for (1S,2S,4R)-(+)-limonene-1,2-diol ( $\geq 97.0\%$ )



All 3 calibration curves have a correlation coefficient close to 1

Good linearity

# Influence of the extraction solvent

## Sample:

- 1.60 M octanoic acid
- 1.6 M  $\alpha$ -phellandrene
- PBS pH 8
- 4.39% (v/v) lipase *Aspergillus niger*
- 2.81% (v/v) hydrolase Re-LEH
- 0.44 mM H<sub>2</sub>O<sub>2</sub>

24 h  
25°C  
1000 rpm  
(Thermoshaker)

## Extraction:

- 1:1 volume ratio in ethyl acetate or n-pentane

- ✓ Non-polar → better for extracting the non-polar substrate
- X Highly volatile

- ✓ Low polarity
- ✓ Less volatile

Agitation at room temperature (30 min)

**GC-MS**

- The difference between the determined concentrations is low
- The conversion is not affected substantially

**Ethyl acetate**

Extraction solvent	Unreacted $\alpha$ -phellandrene (mols/L)	Conversion (%)
Ethyl acetate	$1.25 \times 10^{-1}$	89
n-pentane	$1.67 \times 10^{-1}$	90

# Testing two substrates for the developed process

## 2 samples:

- 1.60 M octanoic acid
- 1.6 M  $\alpha$ -phellandrene or 1.59 M (+)-limonene
- PBS pH 8
- 4.39% (v/v) lipase *Aspergillus niger*
- 2.81% (v/v) hydrolase Re-LEH
- 0.44 mM H<sub>2</sub>O<sub>2</sub>

- Extraction (ethyl acetate)
- Acetylation
- GC-FID



The conversion of  $\alpha$ -phellandrene is influenced by the reaction environment

The difference in selectivity is not reflected in the amount obtained

p-cymene is detected for both cases

Enantiomeric excess around 90 % in both cases

Process performance	$\alpha$ -phellandrene	(+)-limonene
Conversion (%)	87	62
Selectivity for epoxide (%)	0.1	2.0
Epoxide (mols/L)	$1.82 \times 10^{-2}$	$2.37 \times 10^{-2}$
Selectivity for p-cymene (%)	17	27
Selectivity for diols (%)	83	71
Diols (mols/L)	1.25	0.75
Enantiomeric excess (%)	91	87

# Effect of H<sub>2</sub>O<sub>2</sub> on the bienzymatic process

Catalyzes the oxidation of octanoic acid

lipase, H<sub>2</sub>O<sub>2</sub>

-H<sub>2</sub>O

Indirect biocatalysis

Re-LEH, H<sub>2</sub>O

Oxidation agent: H<sub>2</sub>O<sub>2</sub>

$\alpha$ -phellandrene

$\alpha$ -phellandrene epoxide

p-menth-5ene-1,2-diol

Octanoic peracid

H<sub>2</sub>O<sub>2</sub> 30%

1. 0.44 mM H<sub>2</sub>O<sub>2</sub>
2. 0.23 mM H<sub>2</sub>O<sub>2</sub>
3. 0.12 mM H<sub>2</sub>O<sub>2</sub>
4. 0.02 mM H<sub>2</sub>O<sub>2</sub>
5. 0.01 mM H<sub>2</sub>O<sub>2</sub>

H<sub>2</sub>O<sub>2</sub> 30% 1:10 in water

GC-FID

24 h  
25°C  
1000 rpm  
(Thermoshaker)

- 1.60 M octanoic acid
- 1.6 M  $\alpha$ -phellandrene
- PBS pH 8
- 4.39% (v/v) lipase
- 2.81% (v/v) hydrolase
- H<sub>2</sub>O<sub>2</sub>

24 h  
25°C  
1000 rpm  
(Thermoshaker)

**Extraction:**

- 1:1 volume ratio in ethyl acetate

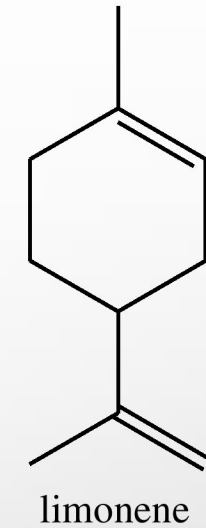
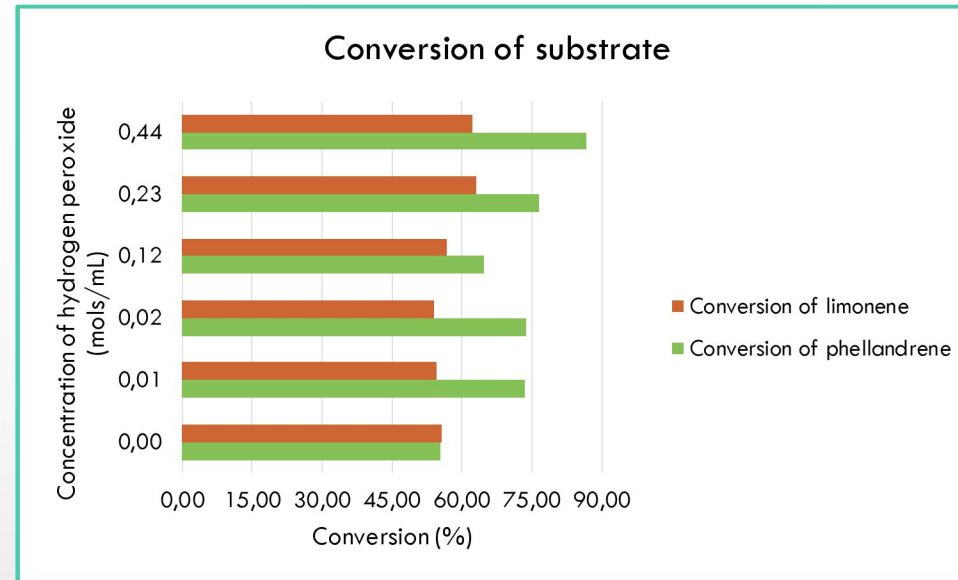
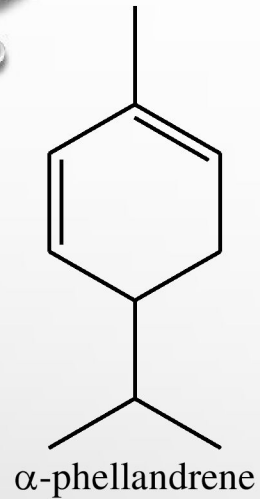
Agitation at room temperature (30 min)

**Acetylation:**

- 100  $\mu$ L sample
- 150  $\mu$ L pyridine
- 66  $\mu$ L acetic anhydride



# Effect of H<sub>2</sub>O<sub>2</sub> on the bienzymatic process



- **Maximum conversion: 87%** for 0.44 mM H<sub>2</sub>O<sub>2</sub>

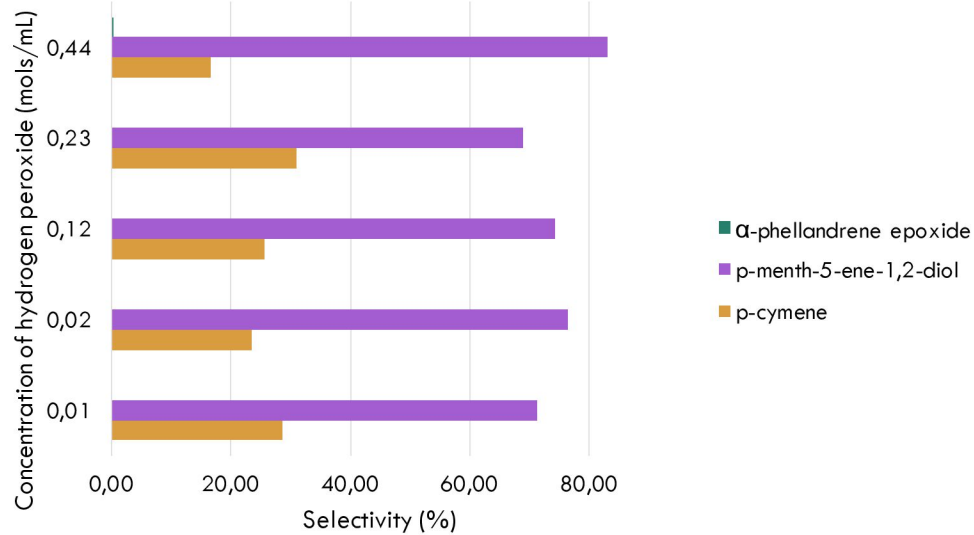
- **Maximum conversion: 63%** for 0.23 mM H<sub>2</sub>O<sub>2</sub>

- The conversion increases together with the concentration of hydrogen peroxide

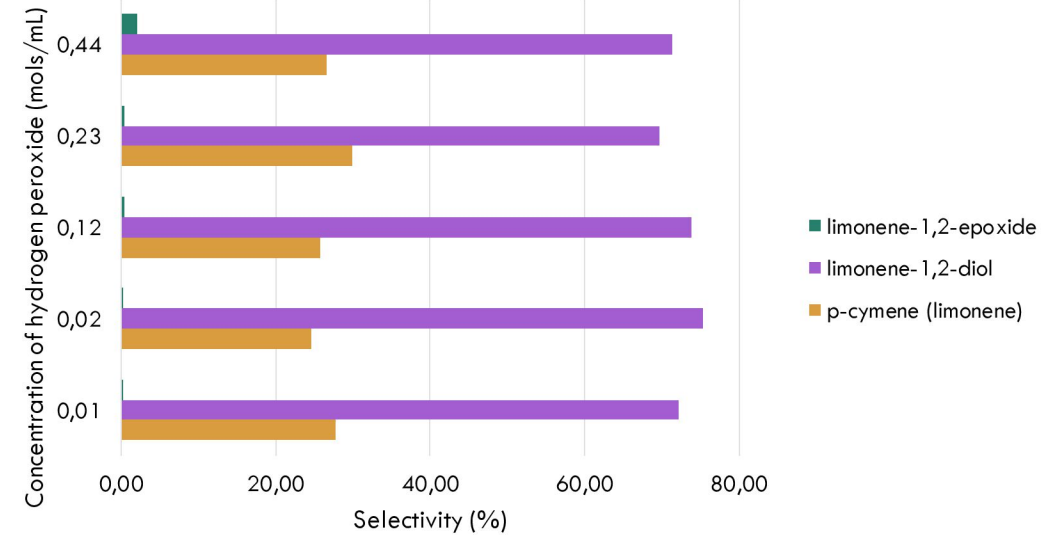
**Better transformation of phellandrene compared with limonene!**

# Effect of H<sub>2</sub>O<sub>2</sub> on the bienzymatic process

Product selectivity for  $\alpha$ -phellandrene substrate



Product selectivity for (R)-(+)-limonene substrate



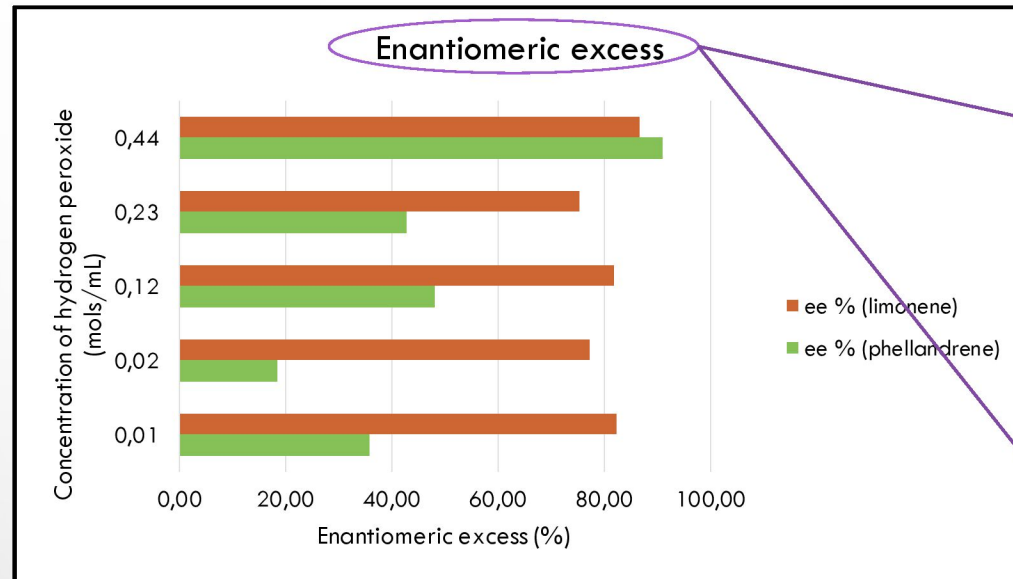
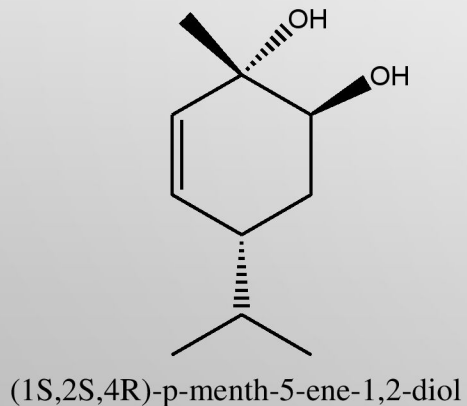
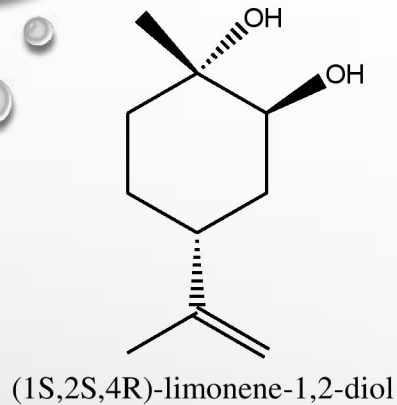
## 0.44 mM H<sub>2</sub>O<sub>2</sub>

- Presence of the epoxide
- the highest diol selectivity (83%)
- the lowest p-cymene selectivity (17%)

- Low selectivity for p-cymene and epoxide intermediates
- Above 70% selectivity for diols

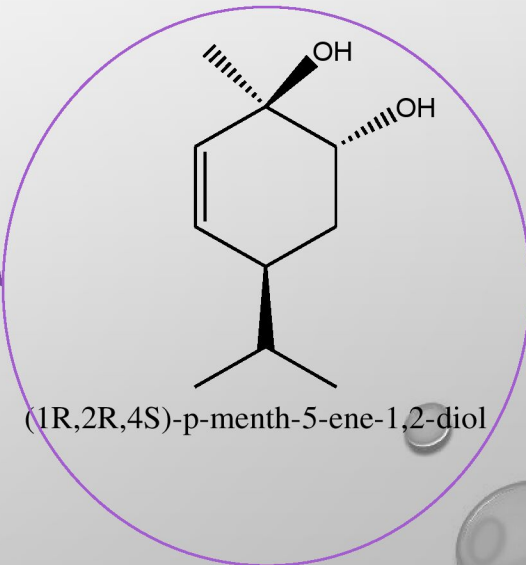
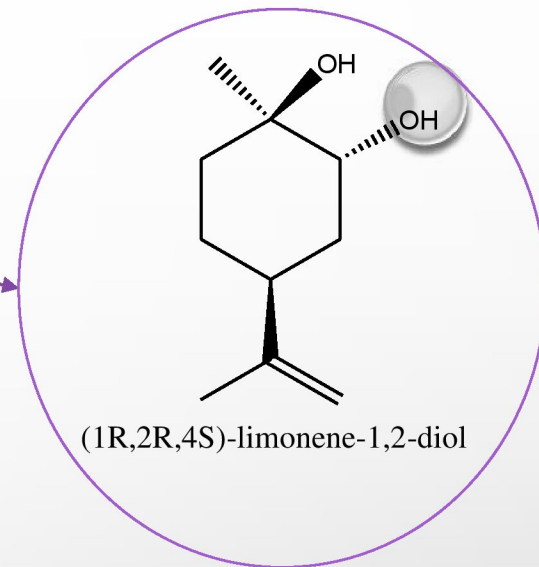
- Epoxide selectivity decreases together with the concentration of H<sub>2</sub>O<sub>2</sub>
- Selectivity for p-cymene between 25 – 30%
- Diol selectivity around 75%

# Effect of H<sub>2</sub>O<sub>2</sub> on the bienzymatic process



$$ee\% = \frac{R - S}{R + S} * 100$$

- Highest ee% for 0.44 mM H<sub>2</sub>O<sub>2</sub> (87% for limonene and 91% for phellandrene)
- ee% decreases at lower H<sub>2</sub>O<sub>2</sub> concentrations
- ee% for α-phellandrene is more affected by H<sub>2</sub>O<sub>2</sub> concentration than limonene



# Efficiency of the bienzymatic system

## Sample 1 (bienzymatic system):

- 1.60 M octanoic acid
- 1.6 M  $\alpha$ -phellandrene
- PBS pH 8
- 4.39% (v/v) lipase *Aspergillus niger*
- 2.81% (v/v) hydrolase Re-LEH
- 0.44 mM  $H_2O_2$

## Sample 2 (mono-enzymatic system):

- 1.60 M octanoic acid
- 1.6 M  $\alpha$ -phellandrene
- PBS pH 8
- 4.39% (v/v) lipase *Aspergillus niger*
- 0.44 mM  $H_2O_2$

## Sample 3 (without biocatalyst):

- 1.60 M octanoic acid
- 1.6 M  $\alpha$ -phellandrene
- PBS pH 8
- 0.44 mM  $H_2O_2$

### Extraction:

- 1:1 volume ratio in ethyl acetate

Agitation at room temperature (30 min)

### Acetylation:

- 100  $\mu$ L sample
- 150  $\mu$ L pyridine
- 66  $\mu$ L acetic anhydride

24 h  
25°C  
1000 rpm  
(Thermoshaker)

GC-FID

24 h  
25°C  
1000 rpm  
(Thermoshaker)

Sample:	Bienzymatic system	Mono-enzymatic system	Without biocatalyst
p-menth-5ene-1,2-diol (mols/L)	1.25	1.41	0.88

The presence of at least one enzyme results in higher diol amount

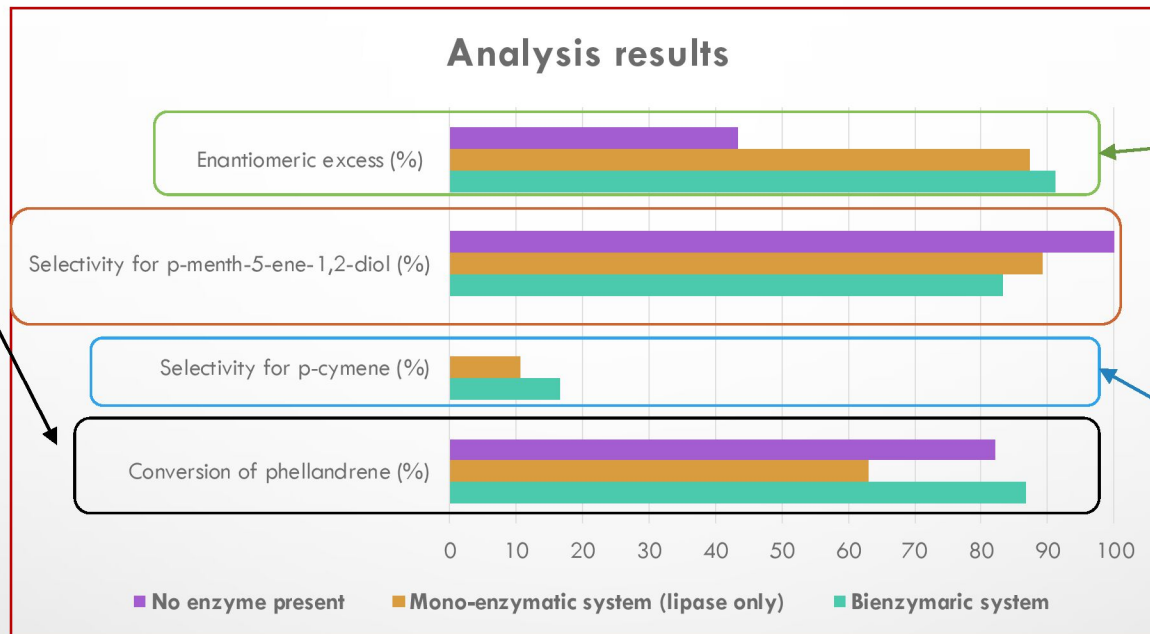
Lowest value

# Efficiency of the bienzymatic system

## Conversion:

- Highest: bienzymatic system (87%)
- Lowest: mono-enzymatic system (63%)

At this moment we do not have an explanation for the high conversion degree obtained in the system without biocatalysts, because we were unable to identify all products.

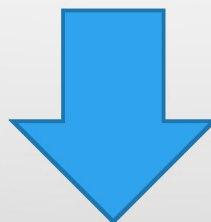


**ee%:**

- Highest: bienzymatic system (91%)
- Lowest: no enzyme system (44%)

**p-cymene selectivity:**

- Highest: bienzymatic system (17%)
- Lowest: no enzyme system (0%)



Sample 1 (bienzymatic system)

Sample 2 (mono-enzymatic system)

Sample 3 (without biocatalysts)

# CONCLUSIONS

- We propose two methods of analysis for our samples, the GC-FID method that can be used for the determination of the enantioselectivity, and GC-MS method for substrate conversion.

- Ethyl acetate is more efficient compared with n-pentane as extraction solvent.

- The biocatalytic system is highly effective in converting both monoterpenoids (phellandrene and limonene) with promising enantioselectivity of the resulted products.

- Hydrogen peroxide concentration has a high influence on the conversion of phellandrene, compared to limonene. However, the highest concentration of hydrogen peroxide offered the best performance of the process.

Optimum reaction mixture:

- 1.6 M octanoic acid
- 1.6 M substrate
- 0.1 M PBS (pH 8)
- 4.39% v/v lipase from *Aspergillus niger*
- 2.81% v/v hydrolase Re-LEH
- 0.44 mM H<sub>2</sub>O<sub>2</sub>

# Future perspectives

1

We propose a more detailed study on the effect of substrate concentration on the conversion, selectivity, and enantiomeric excess.

because

The behavior of the two monoterpenoids used in our study is quite different.

therefore

A study for the substrate influence on the bienzymatic system is requested.

e.g. phellandrene was greatly affected by the concentration of  $H_2O_2$ , but limonene substrate is less influenced by it

2

We propose a biocatalyst designed by immobilization approach of lipase from *Aspergillus niger* and hydrolase Re-LEH.

Why?

Enzyme immobilization is an important step for our design because it provides **higher stability of the biocatalyst**, and the biocatalyst can be **recovered from the reaction phase**, which provides great future perspectives for reusing the biocatalyst.

How?

We propose **cross-linking** and **entrapment**.

Why?

Minimum effect on the catalytic activity of the enzymes

Thank you for your attention!